

Oscillatoria angustissima: A Promising Cu^{2+} Biosorbent

Prerna Ahuja, Rani Gupta, R.K. Saxena

Department of Microbiology, University of Delhi, South Campus, Benito Juarez Road, New Delhi-110021, India

Received: 23 December 1996 / Accepted: 20 February 1997

Abstract. *Oscillatoria angustissima* rapidly adsorbs Cu^{2+} from aqueous solution. The adsorption of Cu^{2+} followed Freundlich Isotherm, and the amount of Cu^{2+} removed from solution increased with increasing Cu^{2+} concentration. The adsorption is pH dependent, and maximum Cu^{2+} removal occurs at pH 5. Of the various pretreatments, HCl treatment of the biomass increased the capacity for Cu^{2+} removal. Presence of Mg^{2+} and Ca^{2+} resulted in decline in the Cu^{2+} adsorption capacity of *Oscillatoria* cells. This species could also effectively remove Cu^{2+} from mine water containing 68.4 $\mu\text{g}/\text{ml}$ of Cu^{2+} at pH 3.45.

Microorganisms can selectively take up various heavy metal ions from aqueous systems and are, therefore, important in regulating environmental pollution.

These organisms remove metals mainly by three phenomena: (i) biosorption [13, 14, 29], (ii) extracellular precipitation [27], and (iii) binding by purified biopolymers [18]. Among these, the use of microbial cells [2, 6] as biosorbents for heavy metals offers a potential and cost-effective alternative to conventional methods for decontamination and/or recovery of heavy metals from a variety of industrial aqueous process streams.

Various types of microbial biomass including both photoautotrophs [7–10] (algae and cyanobacteria) and heterotrophs [2, 4] (bacteria and fungi) have been reported to possess these metal-binding properties.

There are a number of reports on the feasibility of developing technology for removal of metals from polluted water [24] with phototrophs. These phototrophs are generally less resistant/tolerant to heavy metals for growth, but can be grown cheaply on minimal nutritional medium without sugars. Recently, Alga SORB™ (Bio Recovery Systems, Las Cruces, USA), a potential algal biosorbent, has been developed with *Chlorella vulgaris* for waste water treatment. This could efficiently adsorb metals leading to decontamination [1].

The present work deals with copper biosorption by a filamentous, non-heterocystous cyanobacterium, *Oscillatoria angustissima*. The biosorption capacities of treated and untreated biomass have been compared. The poten-

tial for its use in decontamination of mine effluent sample has also been examined.

Materials and Methods

Organism and growth conditions. *Oscillatoria angustissima* culture was obtained from National Facility for Blue Green Algal Collections, (IARI, New Delhi, India). It was grown at $25^\circ \pm 2^\circ\text{C}$ under 1100 lux light intensity in BG11 minimal medium [22]. The medium composition (g/L) is NaNO_3 1.5, K_2HPO_4 0.04, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 0.075, $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ 0.036, citric acid 0.006, ferric ammonium citrate 0.006, EDTA (disodium salt) 0.001, Na_2CO_3 0.02, and trace metal mix 1 ml/L. The composition of trace metal mix in g/L is H_3BO_3 2.86, $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$ 1.81, $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ 0.222, $\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ 0.39, $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ 0.079, $\text{Cu}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ 0.0494.

The pH of the medium was adjusted to 7 ± 0.2 . The filaments were harvested in exponential phase after 10 days of growth by centrifugation at 6000 rpm for 10 min. Thereafter, the filaments were washed thoroughly with deionized distilled water and then used for metal uptake experiments.

Dry weight determination. The dry weight of cells was determined by pelleting a known volume of cell suspension and drying the pellet at 80°C for 48 h until a constant weight was obtained.

Preparation of treated cells. (a) *High temperature treatment.* The cells equivalent to 4 mg dry wt were incubated at 60°C and 100°C in a water bath for 10 min and subsequently washed thoroughly by deionized water and collected by centrifugation.

(b) *Dilute alkali/acid treatment.* The cells equivalent to 4 mg dry wt were treated with 0.01 N NaOH for 10 min and subsequently centrifuged, repeatedly washed, and collected. Similarly, the cells were treated with 0.01 N HCl and used for metal adsorption studies.

All the chemicals used in media preparation and the metal salt ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) were of analytical grade (BDH), and the HCl used was supplied by Merck (Mumbai, India). The standard solution of Cu^{2+} (1000 ppm) for atomic absorption measurements were obtained from National Physical Laboratory, New Delhi, India.

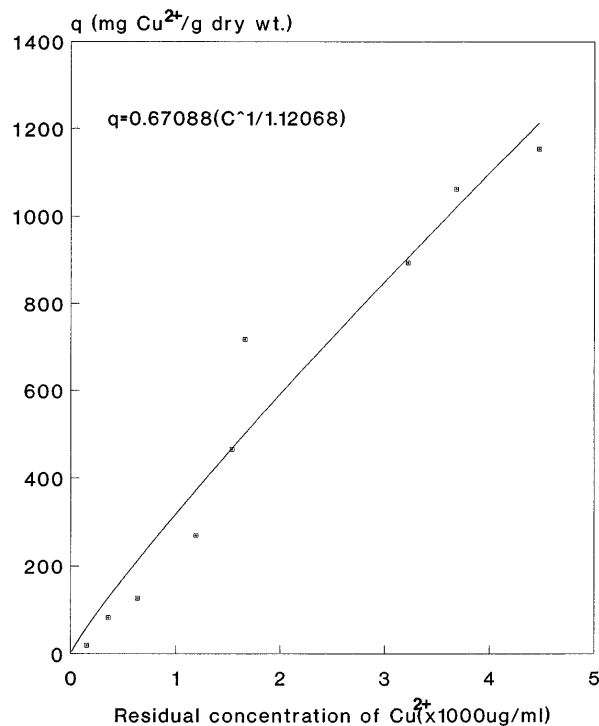


Fig. 1. Freundlich adsorption isotherm for copper at 0.08 mg dry wt/ml biomass concentration (pH 5.0).

Analytical methods. The desired concentrations of Cu^{2+} (5 mg/L to 200 mg/L) were prepared by dissolving $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ in deionized distilled water. Short-term metal uptake experiments were performed in 250-ml Erlenmeyer flasks containing 50 ml of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ solution of known initial concentration (pH 2–5) at 25°C for 2 h unless otherwise mentioned. In all experiments except for the effect of cell density, the biomass concentration was kept constant to 0.08 mg dry wt/ml. Metal-free and biomass-free blanks were used as controls. Biomass-free blanks were used to estimate the exact initial concentration (at 50 mg/L the exact initial concentration as estimated by Atomic Absorption measurements, Shimadzu AA-260 ranged from 48 to 51 mg/L) of Cu^{2+} by dilution. Separation of biomass from metal-bearing solution was achieved through centrifugation at 6000 rpm for 10 min in Remi RC30 centrifuge. The supernatant was appropriately diluted and read for the remaining Cu^{2+} content at 324.5 nm wavelength (Slit width 3.8Å). All experiments were carried out in triplicate and repeated three times.

The metal uptake capacity in mg/g (q) was calculated from the initial concentration (C_i) and the final concentration (C_f) of the metal according to the following equation [28]:

$$q = V(C_i - C_f)/M$$

where V is the liquid sample volume and M is the biomass dry weight. The biosorptive metal uptake was evaluated and expressed by use of Freundlich adsorption model [5].

The general form of model is

$$q = kC^{1/n}$$

This can be linearized by taking natural logarithm in the form of $\ln q = \ln k + 1/n \ln C$.

where q = uptake of species (metal) and C = equilibrium (final/residual) concentration. The intercept $\ln k$ gives a measure of adsorbent capacity and the slope $1/n$ gives the intensity of adsorption.

Results and Discussion

Oscillatoria sp. showed excellent copper biosorption (Figs. 1, 2), and the adsorption behavior of cells could be explained by the Freundlich isotherm model. A log-log plot between the amount of Cu^{2+} in the solution and in the biomass followed a straight line with acceptable correlation coefficient (r^2). All previous studies of adsorption have been evaluated in terms of either Langmuir or Freundlich isotherms. The validity of the model was tested at the biomass concentrations viz biomass equivalent to 0.08 and 0.16 mg dry wt/ml. The high value of r^2 , viz. 0.951 and 0.959 respectively, implies that the curve fits the data fairly well. The application of “t” test showed n to be highly significant for both biomass concentrations, but k was not significant.

Oscillatoria sp. could adsorb Cu^{2+} even at lower concentrations, indicating a good affinity for the metal. The uptake was found to increase with the increase in concentration of Cu^{2+} . It could adsorb about 268.45 mg/g dry wt of Cu^{2+} at a residual concentration of 23 mg/L, and it increased to about 1 g/g Cu^{2+} adsorption at a residual concentration of 89.33 mg/L. *Oscillatoria* sp. showed a high sequestration of copper at lower equilibrium concentrations, as indicated by the steeper isotherm shown in Fig. 2, suggesting a strong binding for copper. Such high capacities for metal adsorption have been reported for few organisms [9, 16].

The effect of increasing concentration of biomass on Cu^{2+} adsorption revealed that, although the amount of Cu^{2+} adsorbed per unit dry weight decreased, the total amount of Cu^{2+} adsorbed increased. *Oscillatoria* sp. could adsorb about 685 mg/g dry wt of Cu^{2+} at 0.04 mg dry wt/ml of biomass, whereas the value decreased to 165 mg dry wt/ml at 0.2 mg dry wt/ml biomass density, but the percentage removal was greater in the latter (Table 1). This confirms the previous observations of adsorption in algae by Horikoshi et al. [10, 11] and Nakajima et al. [17]. Similar results have been obtained in a recent study [26] with *Arthrobacter* sp.; reduction in biomass concentration was shown to increase the specific uptake of copper. The dependence of adsorption on the cell density may be due to electrostatic interactions as more cations are adsorbed on the cell when the cell distances are great, as suggested by Itoh et al. [12].

The time course of uptake of Cu^{2+} showed the adsorption to follow fast kinetics. It was found that 90% of adsorption was complete within the first 15 min of initial contact with the metal-bearing solution.

Adsorption of copper was found to increase with increase in temperature from 25°C to 45°C. *Oscillatoria* sp. showed maximum Cu^{2+} adsorption at 45°C, 427.5

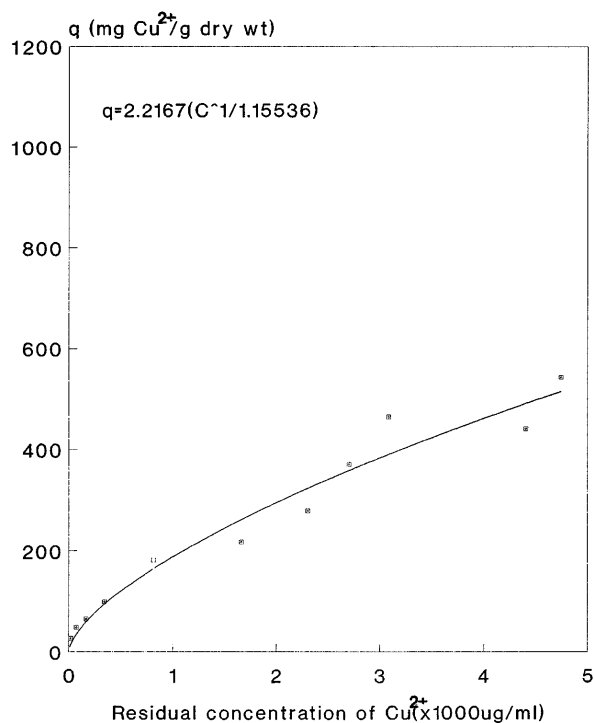


Fig. 2. Freundlich adsorption isotherm for copper at 0.16 mg dry wt/ml biomass concentration (pH 5.0).

Table 1. Cu²⁺ biosorption at varying concentration of biomass by *Oscillatoria* sp.

Biomass concentration (mg dry wt/ml)	mg Cu ²⁺ /g dry wt
0.04	685.0 ± 45.00
0.08	352.5 ± 13.20
0.12	234.9 ± 9.20
0.20	165.0 ± 8.70

pH 5.0; temperature, 25°C.

mg/g dry wt of Cu²⁺, and minimum adsorption at 25°C. de Rome and Gadd [4] have also reported temperature-dependent Cu²⁺ adsorption in filamentous fungi. Higher uptake capacities at increased temperature have also been reported for uranium by Shumate et al. [21], Strandberg et al. [23], and Tsezos and Volesky [25].

The adsorption of Cu²⁺ was strongly affected by pH. The adsorption capacity of *Oscillatoria* sp. increased with increasing pH, to a maximum at pH 5.0 (327 ± 10.60 mg/g dry wt). Kuyucak and Volesky [14] also have similar reports in *Sargassum natans*, where they have found the optimum pH for biosorption of Cu²⁺ to be 4–5. The experiments could not be conducted at higher pH values because of precipitation of copper hydroxide above pH 5.0.

At very low pH there is an increase in metal availability and mobility, but protons tend to compete [19] with the cations to bind to the sites, thus lowering the extent of biosorption.

The presence of some competing ions (particularly those present in hard water) generally affects the adsorption of heavy metal ions and can reduce the efficiency of metal removal [20]. The various groups implicated as participating in divalent metal ion binding according to Christ et al. [3] include the carboxylate, amine, imidazole, phosphate, sulfate, sulfhydryl, and other functional groups in cell surface proteins and sugars.

In the present study the divalent cations, viz. Mg²⁺ and Ca²⁺, were found to inhibit Cu²⁺ adsorption. The Cu²⁺ adsorption was not significantly affected by 100 mg/L of Ca²⁺, but doubling of Ca²⁺ concentration resulted in 21.2% decline in Cu²⁺ adsorption capacity of *Oscillatoria* sp. Similar results were obtained with Mg²⁺ ions, but the percentage decline in Cu²⁺ adsorption was slightly more at 200 mg/L Mg²⁺ (30.06%) than with Ca²⁺. The inhibition by 100 mg/L Mg²⁺ was found to be about 12%.

Pretreatment of biomass either by physical or chemical treatments [20] or crosslinking of biomass is known to alter the biosorption capacity of biomass [15]. Biomass of *Oscillatoria* sp. was differentially treated to observe the change in potency of biosorption for Cu²⁺. Of the various treatments, only washing the biomass with 0.01 N HCl resulted in increased Cu²⁺ adsorption to about 420 mg/g dry wt compared with 385 mg/g dry wt of the native biomass. Sampedro et al. [20] have reported an increased biosorption in a filamentous cyanobacterium *Phormidium laminosum* after NaOH treatment, but HCl treatment was ineffective. The observed results clearly indicate that some pretreatments do expose more metal-binding groups either by removing some of the masking groups or by a change in configuration [19]. Also, since the dried (dead) biomass could sequester copper to the same extent, the absence of an active energy-dependent mechanism is suggested.

Oscillatoria angustissima was investigated for its Cu²⁺ removal ability from effluent sample of Ghatsila Copper Mines (Bihar, India) containing about 68.4 mg/L Cu²⁺ at pH 3.45. It was found that native biomass could adsorb about 202.5 mg/g dry wt. of Cu²⁺ from mine water in batch experiments, and the total capacity for its Cu²⁺ removal increased with doubling the biomass concentration, but the mg/g removal decreased to about 118.75.

The rapid, extremely high capacity of *Oscillatoria* sp. for Cu biosorption and removal of copper from mine effluents makes this fresh water cyanobacterium a promising future biosorbent. Further work is in progress in the

direction of formulating *Oscillatoria* sp. to an economical biosorbent material and to achieve maximum desorption of Cu^{2+} from the biomass for its repeated use in subsequent cycles.

ACKNOWLEDGMENTS

The present work was financially supported by the University Grants Commission. Thanks are due to Prof. Daljit Singh for his valuable help in statistical analysis and preparation of isotherms.

Literature Cited

1. Bedell GW, Darnall DW (1990) Immobilization of non viable, biosorbent, algal biomass for the recovery of metal ions. In: Volesky B (ed) Biosorption of heavy metals. Boca Raton: CRC Press, pp 313–326
2. Chang Jo-S, Hong J (1994) Biosorption of mercury by the inactivated cells of *Pseudomonas aeruginosa* PU21 (Rip64). Biotechnol Bioeng 44:999–1006
3. Christ RH, Obserholser K, Shank N, Nguyen M (1981) Nature of bonding between metallic ions and algae cell walls. Environ Sci Technol 15:1212–1217
4. deRome L, Gadd GM (1987) Copper adsorption by *Rhizopus arrhizus*, *Cladosporium resinae* and *Penicillium italicum*. Appl Microbiol Biotechnol 26:84–90
5. Freundlich H (1926) Colloid and capillary chemistry. London: Methuen
6. Gadd GM (1990) Heavy metal accumulation by bacteria and other microorganisms. Experientia 46:834–840
7. Gale NL, Wixon BG (1979) Removal of heavy metals from industrial effluents by algae. Dev Ind Microbiol 20:259–273
8. Greene B, Darnall DW (1990) Microbial oxygenic photoautotrophs (cyanobacteria and algae) for metal ion binding. In: Ehrlich HL, Brierley C (eds) Microbial metal recovery. New York: McGraw Hill pp 277–302
9. Holan ZR, Volesky B (1994) Biosorption of lead and nickel by biomass of marine algae. Biotechnol Bioeng 43:1001–1009
10. HoriKoshi T, Nakajima A, Sakaguchi T (1979) Uptake of uranium from sea water by *Synechococcus elongatus*. J Ferment Technol 57:191–194
11. Horikoshi T, Nakajima A, Sakaguchi T (1981) Studies on the accumulation of heavy metal elements in biological systems XIX. Accumulation of uranium by microorganisms. Eur J Appl Microbiol Biotechnol 12:90–96
12. Itoh M, Yuasa M, Kobayashi T (1975) Adsorption of metal ions on yeast cells at varied cell concentrations. Plant Cell Physiol 16:1167–1169
13. Kuyucak N, Volesky B (1988) Biosorbents for recovery of metals from industrial solutions. Biotechnol Lett 10:137–142
14. Kuyucak N, Volesky B (1989) Accumulation of cobalt by marine alga. Biotechnol Bioeng 33:809–814
15. Leusch A, Holan ZR, Volesky B (1995) Biosorption of heavy metals (Cd, Cu, Ni, Pb, Zn) by chemically-reinforced biomass of marine algae. J Chem Technol Biotechnol 62:279–288
16. Luef E, Prey T, Kubicek, CP (1991) Biosorption of zinc by fungal mycelial wastes. Appl Microbiol Biotechnol 34:688–692
17. Nakajima A, Horikoshi T, Sakaguchi T (1981) Studies on the accumulation of heavy metal elements in biological systems. XVII. Selective accumulation of heavy metal ions by *Chlorella regularis*. Eur J Appl Microbiol Biotechnol 12:76–83
18. Norberg B, Persson H (1984) Accumulation of heavy metal ions by *Zoogloea ramigera*. Biotechnol Bioeng 26:239–246
19. Paknikar KM, Palnitkar US, Puranik PR (1993) Biosorption of metals from solution by mycelial waste of *Penicillium chrysogenum*. In: Torma AE, Apel ML, Brierley CL (eds) Biohydrometallurgical technologies, Vol. II. The Minerals, Metals & Materials Society. Warrendale, Pennsylvania: TMS Publication, pp 229–235
20. Sampedro MA, Blanco A, Llama MJ, Serra JL (1995) Sorption of heavy metals to *Phormidium laminosum* biomass. Biotechnol Appl Biochem 22:355–366
21. Shumate SE, Strandberg GW, Parrott (1978) Biological removal of metal ions from aqueous streams. Biotechnol Bioeng 8:13–20
22. Stanier RY, Kunisawa R, Mandel M, Cohen Bazire G (1971) Purification and properties of unicellular blue green algae (order Chroococcales). Bacteriol Rev 35:171–205
23. Strandberg GW, Shumate SE, Parrott JR (1981) Microbial cells as biosorbents for heavy metals: accumulation of uranium by *Saccharomyces cerevisiae* and *Pseudomonas aeruginosa*. Appl Environ Microbiol 41:237–245
24. Ting YP, Lawson F, Prince IG (1989) Uptake of cadmium and zinc by the alga *Chlorella vulgaris*: Part I. Individual ion species. Biotechnol Bioeng 34:990–999
25. Tsezos M, Volesky B (1981) Biosorption of uranium and thorium. Biotechnol Bioeng 23:583–604
26. Veglio F, Beolchini, F, Gasbarro A (1997) Biosorption of toxic metals: an equilibrium study using free cells of *Arthrobacter* sp. Process Biochem 32:99–105
27. Volesky B (1987) Biosorbents for metal recovery. Trends Biotechnol 5:96–101
28. Volesky B (1990) Removal and recovery of heavy metals by biosorption. In: Volesky B (ed) Biosorption of heavy metals. Boca Raton: CRC Press, pp 8–43
29. Volesky B (1994) Advances in biosorption of metals: selection of biomass types. FEMS Microbiol Rev 14:291–302